

Figure 1: Metastatic dissemination and tumour microenvironment. In addition to the sprouting of neighbouring pre-existing vessels, tumoural angiogenesis is supported by the mobilization of different cell types including hematopoietic stem cells (HSC), endothelial progenitor cells (EPC) and mesenchymal stem cells (MSC). Lymphangiogenesis facilitates metastasis by providing an alternative route of dissemination. Primary tumoural cells could induce the establishment of a pre-metastatic niche preceding metastatic colonization of the targeted organ. CSC = cancer stem cells

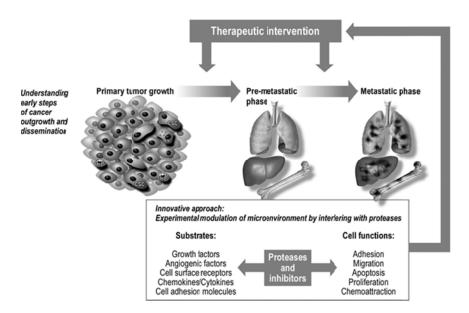


Figure 2: Aims and approaches used by the *MEM consortium*. The study provides new insights into the interplay between cancer cells, cancer stem cells (CSC) and their microenvironment, both at primary and secondary sites. Modulation of tumour microenvironment has been achieved by interfering with proteases that are central mediators of a complex molecular network, as well as key regulators of various cell functions.

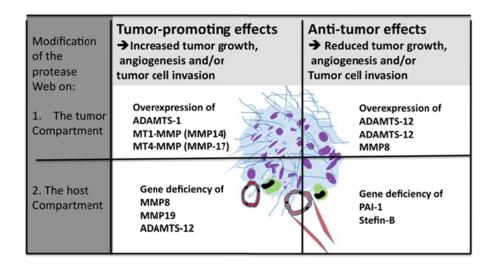


Table 1: summary of the tumour-promoting effects and the anti-tumoural actions of proteases and their inhibitors demonstrated by the MEM consortium.

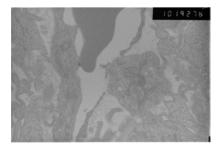


Figure 3: Illustration of the intravasation of a tumour cell frequently observed in MT4-MMP overexpressing tumours, but not in control tumours.

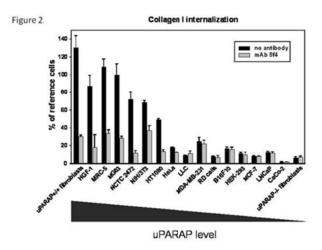


Figure 4: uPARAP/Endo180 mediates collagen internalization in a variety of cell types. Collagen internalization is inhibited by mAb 5f4 for a variety of established cell lines. Collagen internalization was measured for each cell line. Internalization was carried out in the absence of antibody (black columns) or in the presence of 10 μ g/ml of mAb 5f4 (grey columns). The cell lines are ranked in the diagram according to their level of uPARAP/Endo180 expression, with the highest expressing cells to the left.

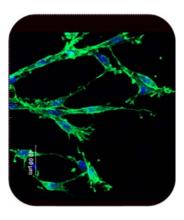


Figure 5: Illustration of lymphatic endothelial cells spreading from a lymphatic vessel fragment. Endothelial cells are labeled in green (with an anti-LYVE-1 antibody) and nuclei are stained in blue. This model reproduces the different steps of the lymphangiogenic process observed during cancer progression, both at primary site and secondary sites of tumour growth.

Kinetics of tumor growth and lung metastasis in the

MMTV - PyMT mouse model for breast cancer Primary tumor growth First palpable **Tumor size** p50 p80 p60 p0 p30 p70 p98 Time points of material collection for miRNA profiling Normal Adenosis/Hyperproliferation Mammary gland Ductal Carcinoma in situ histology **Ductal Adenocarcinoma** No Mets = premetastatic phase Micrometastases Lung metastasis (based on histology Metastases

Figure 6: Kinetics of tumor growth and metastasis formation.

and PyMT expression in the lung)

Table 1 Tumor-associated immune cells

Cell type	Marker	PymT+10;wt % of total ± s.e.m.	PymT+10;Tg(CTSB)+10 % of total±s.e.m.	
CD4 + T cells	CD4	0.60±0.17	0.61 ± 0.23	NS
CD8 + T cells	CD8	1.05 ± 0.15	1.79 ± 0.47	NS
B cells	CD19	0.51 ± 0.07	1.47 ± 0.55	P<0.05
Macrophages	F4,80	2.00 ± 0.30	2.83 ± 0.66	NS
Neutrophils	7/4 Antigen	1.77 ± 0.41	2.25 ± 0.75	NS
Mast cells	CAE	18.8 ± 3.1 h	31.7 ± 1.5°	P<0.01

Quantification of distinct immune cell types, including T and B cells, macrophages and neutrophils as percentage of total cells by flow cytometry (n = 8-12 per genotype). Mast cells were identified by CAE histochemistry and the number of CAE+ cells per mm² of tumor stroma was calculated from three independent sectional planes (a = 4 per group). Data are presented as means and standard errors; statistical analysis was done by Student's t-test.

"Detected by anti-mouse neutrophils/Clone 7/4; rat IgG2a (Cedarlane, Burlington, Ontario, Canada).
"Number of mast cells per mm" tumor stroma.

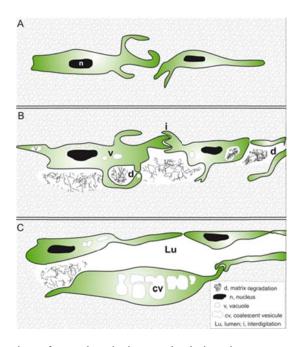


Figure 7: Model of formation of new lymphatic vessels during the process of lymphangiogenesis (see description in the text).

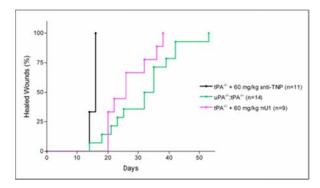


Figure 8: mU1 retards wound healing, demonstrating the efficient blocking of uPA function in a physiological invasive process in vivo.

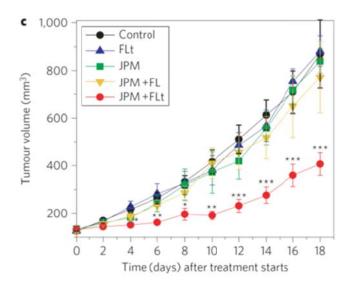


Figure 9: Anti-tumour effect of magnetically targeted ferri-liposomes containing JPM-565. Tumour volumes for each treatment day for the different treatment groups. Mice were treated with ferri-liposomes without (FL) and with magnetic targeting (FLt), and JPM-565 (JPM) combinations as represented by the '+' sign.