

Figure 1. Zebrafish activity in response to temperature change.

Zebrafish locomotor (swimming) activity in response to temperature change was quantified by using a Zebrafish system (Viewpoint). Before the temperature interval was started, a baseline of zebrafish activity was recorded for 10 min. During this baseline measurement the temperature of the water is 28.5°C (black line). After the start of the temperature interval the maximum temperature of 35°C is reached within 3-4 minutes. (A) Overview of the activity of WT zebrafish (Blue line) and the mutant *scn8aa* Loss of function (LOF) line (orange) in response to temperature change (black line). (B) Quantification of the activity of WT (blue bar) and *scn8aa* LOF mutant zebrafish (orange bar) for a period of 5 minutes from the moment when the maximum temperature is reached. Statistical analysis was performed using a Student's *t*-test ($P \leq 0.01$)

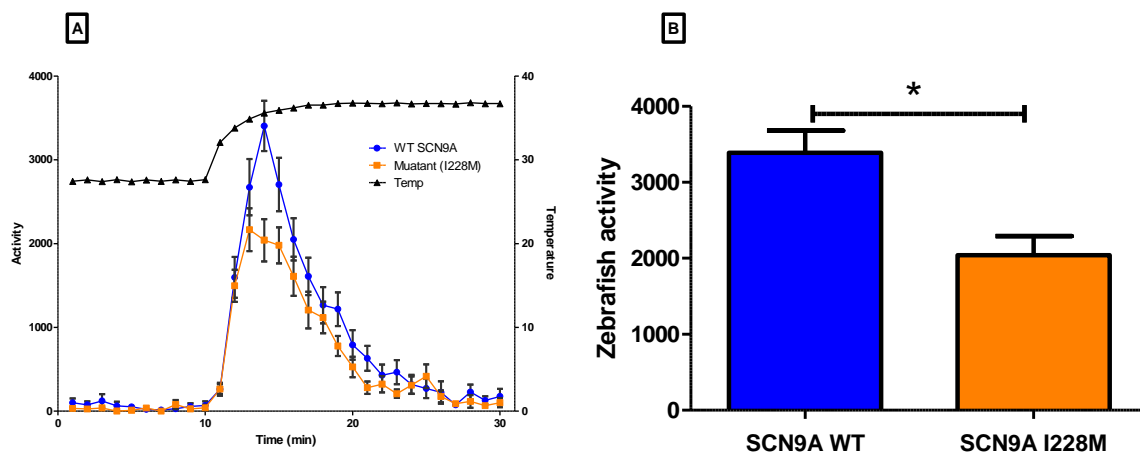


Figure 2. Zebrafish activity in response to temperature change.

Zebrafish activity @ 4dpf in response to temperature change was quantified by using the Zebrafish system (Viewpoint). Before the temperature interval was started, a baseline of zebrafish activity was recorded for 10 min. During this baseline measurement the temperature of the water is 28.5°C (black line). After the start of the temperature interval the maximum temperature of 36°C is reached within 3-4 minutes. (A) Overview of the activity of WT zebrafish (Blue line) and the mutant (I228M) line (orange) in response to temperature change (black line). (B) Quantification of the activity of WT (blue bar) and mutant zebrafish (orange bar) at the maximum temperature. Statistical analysis was performed with a Student's *t*-test ($P \leq 0.05$)

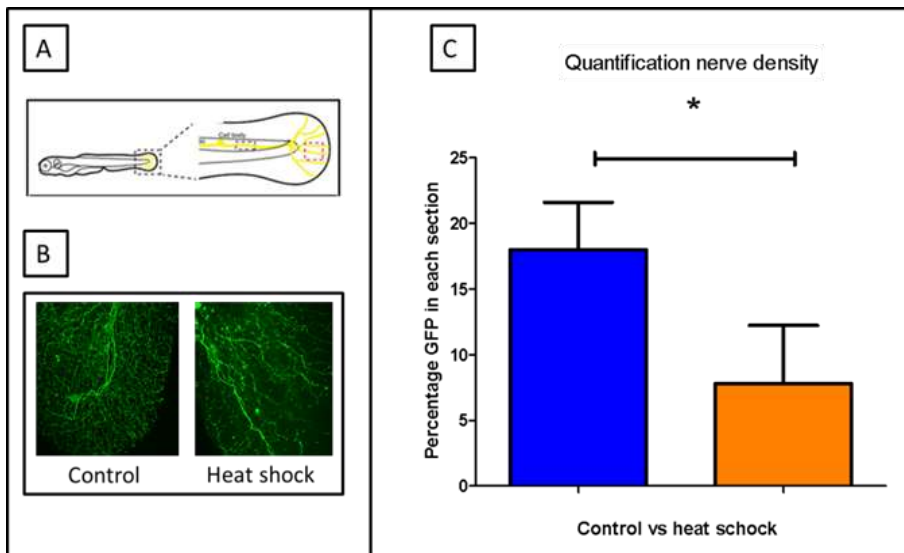


Figure 3: Quantification of the density of sensory neurons in zebrafish embryos after heat shock.

The tail section of the zebrafish (A) is a very thin section and gives a good representation of the nerve fibers in the zebrafish (Campbell, P. D. et al. *J. Neurosci.* 34, 14717–14732 (2014)). After a heat shock of 5 seconds at 42°C the nerve density of the zebrafish is significantly reduced (B and C). Quantification is performed with the ImageJ plugin particle analyzer after thresholding. For each recording the percentage of GFP was calculated in 3 at random areas (C).

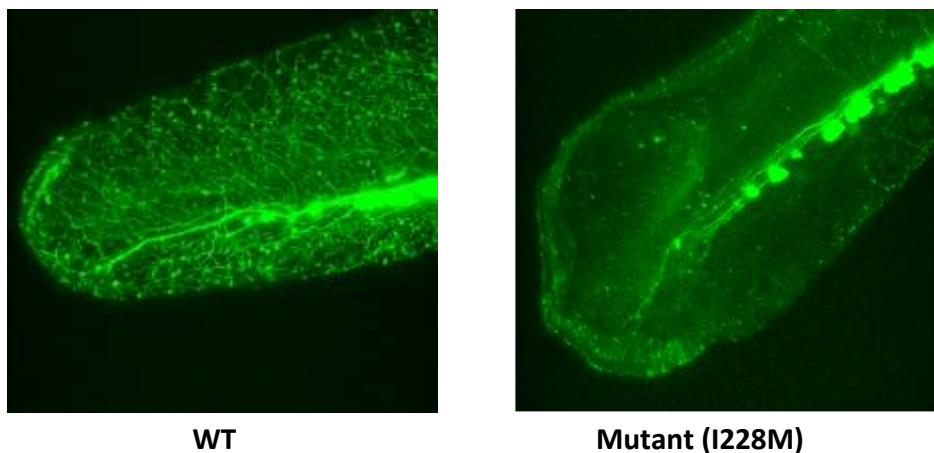


Figure 4. Confocal recordings WT (left panel) vs I228M-Mutant (right panel)

Confocal recordings of the tail section of WT (left) and Mutant (right) 2-day-old zebrafish; maximal projection of Z-stack.

Table 1: Functional assessment of variants in voltage-gated sodium channels

| Mutation | Voltage-clamp | Current-clamp | Reference |
|---------------|------------------|------------------------|------------------|
| Nav1.6 | | | |
| Leu1284Gln | Not Done | No Change | unpublished |
| Nav1.7 | | | |
| Met130Ile | No Change | Not Done | unpublished |
| Lys665Arg | No Change | Not Done | unpublished |
| Val810Met | Not Done | No Change | unpublished |
| Thr920Asn | No Change | Not Done | unpublished |
| Met1230Thr | No Change | Not Done | unpublished |
| Asn1245Ser | No Change | Not Done | unpublished |
| Met1852Thr | Gain-of-Function | Not Done | Pending |
| Nav1.8 | | | |
| Pro721Leu | Gain-of-Function | Pending | Unpublished |
| Arg1582His | Loss-of-function | Increased Excitability | Unpublished |
| Nav1.9 | | | |
| Ile381Thr | Gain-of-Function | Increased Excitability | Huang et al 2014 |
| Ala681Asp | No current | Not Done | Unpublished |
| Lys682Ile | Not Done | No Change | Unpublished |
| Gly699Arg | Gain-of-Function | Increased Excitability | Han et al 2015 |
| Ile807Val | Gain-of-Function | Pending | Unpublished |
| Ala864Pro | No Major change | No Major Change | Unpublished |
| Leu1158Pro | Gain-of-Function | Increased Excitability | Huang et al 2014 |
| Val1437Met | Loss-of-Function | Not Done | Unpublished |
| Phe1689Leu | Mixed Phenotype | pending | Unpublished |
| Nav2 | | | |
| Asp109Asn | Pending | Increased Excitability | Unpublished |

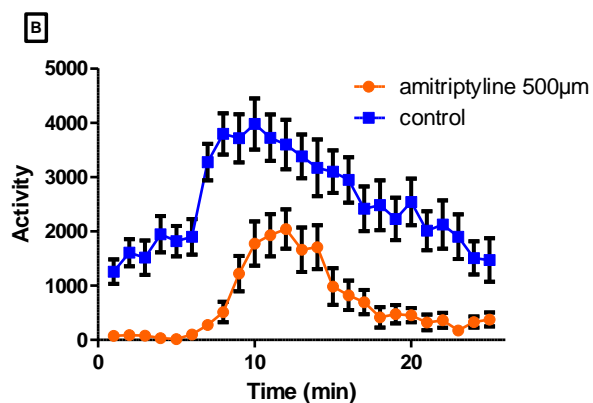
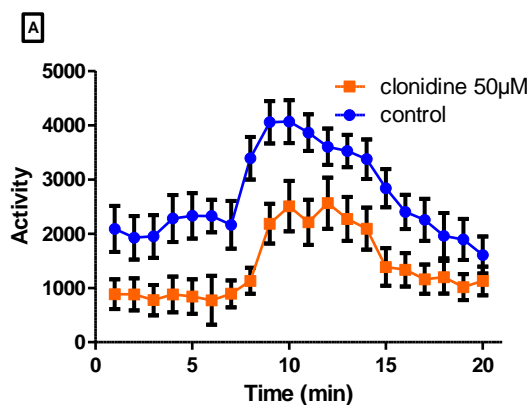


Figure 5. The effect of high concentrations of clonidine and amitriptyline on zebrafish activity in response to noxious temperatures.

The left panel represents the activity of 5-day-old zebrafish larvae in response to a temperature increase starting at 6 minutes and reaching a maximum of 36.5 °C. During these experiments, zebrafish larvae were incubated in Clonidine 50µM (A) and Amitriptyline 500µM (B).

| Compound | Concentration |
|-----------------|----------------------|
| Carbamazepine | 121µM |
| Lamotrigine | 150µM |
| Amitriptyline | 0.5µM |
| Clonidine | 5µM |

Table 2. Used concentrations of the compounds

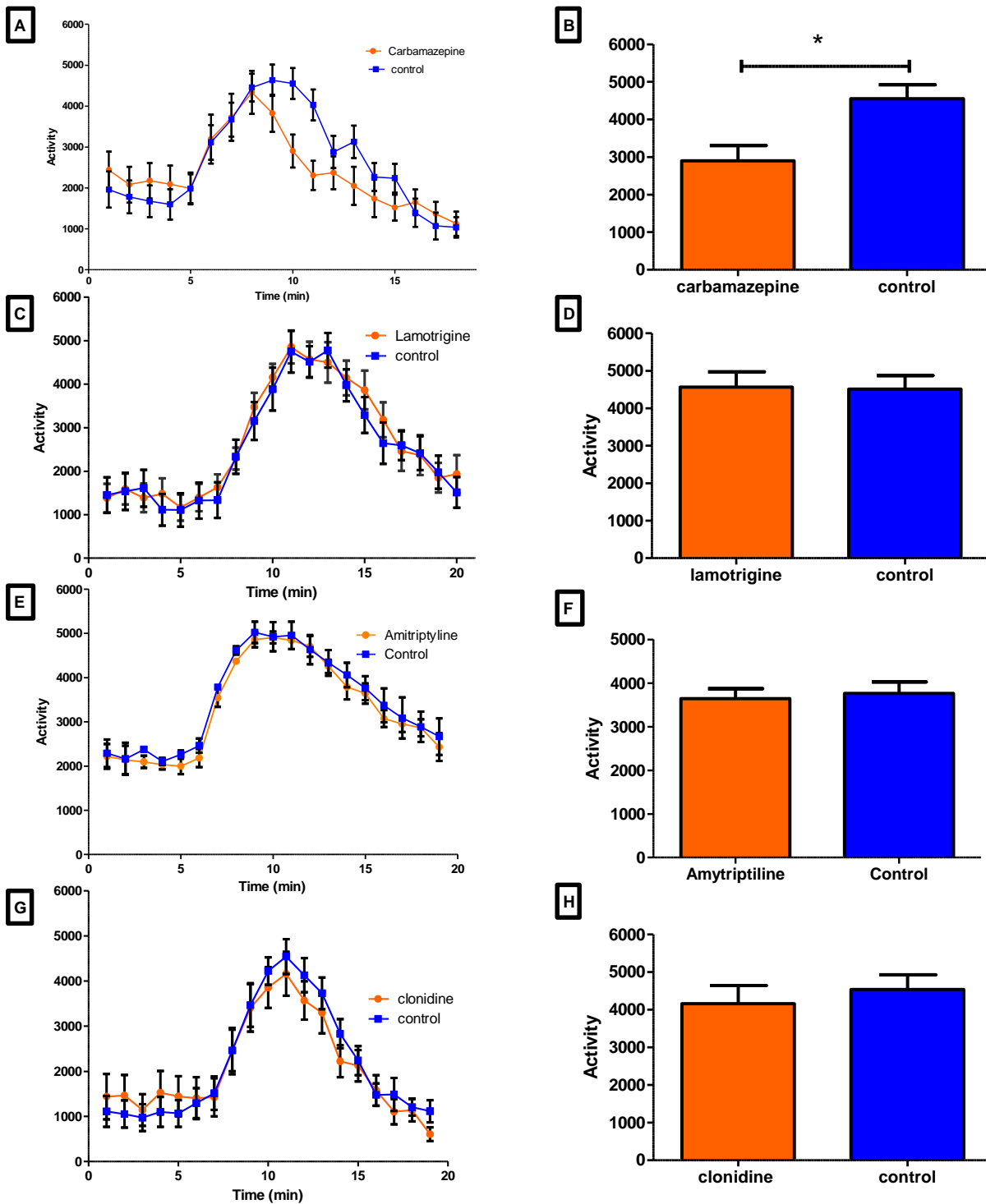


Figure 6. The effects of pain medication on zebrafish activity in response to noxious temperatures.

The left panel represents the activity of zebrafish in response to a temperature increase starting after 6 minutes and reaching a maximum of 36.5 °C. The right panel presents the activity of 5-day-old zebrafish at the maximal temperature (36.5 °C). The zebrafish were during the experiments incubated in carbamazepine (A,B), Lamotrigine (C,D), Amitriptyline (E,F) and Clonidine (G,H).

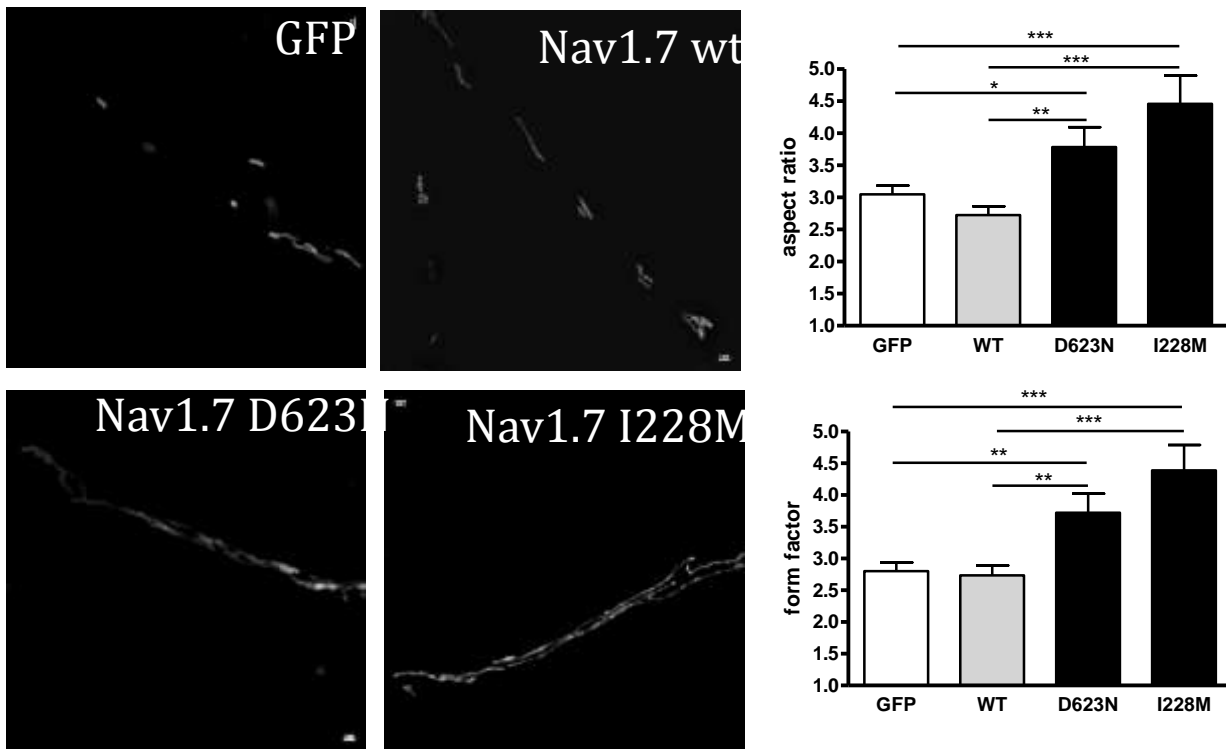


Figure 7. The effects of NaV1.7 mutation on mitochondrial shape.

Quantitative analysis of mitochondrial morphology in rat DRG neurons co-transfected with pDsRed2-Mito and GFP plasmid, Nav1.7 wt or mutant D623N and I228M. Representative confocal images show mitochondrial shape. The histograms show AR and FF media values for all mitochondria (>300 mitochondria particles, 10 cells analyzed per condition).